

Fish *Balistes capriscus* skin extract-induced relaxation in mesenteric arterial bed of rat

L.S. Cavalli^{a,b}, P.L. Possette^a, B. Schmidt^a, C. Kruel^a, M. Grando^a, E. Badiale Furlong^c,
M.R. Cezar-Vaz^{d,e}, D.M. Barros^a, A.L. Muccillo-Baisch^{a,b,e,*}

^a Departamento de Ciências Fisiológicas, Universidade Federal do Rio Grande, Rio Grande, Rua Eng. Alfredo Huch 475, Rio Grande, 96201-900, Brazil

^b Programa de Pós-Graduação em Fisiologia Animal Comparada, Setor de Farmacologia, Universidade Federal do Rio Grande,
Rio Grande, Rua Eng. Alfredo Huch 475, Rio Grande, 96201-900, Brazil

^c Departamento de Química, Universidade Federal do Rio Grande, Rio Grande, Rua Eng. Alfredo Huch 475, Rio Grande, 96201-900, Brazil

^d Departamento de Enfermagem, Universidade Federal do Rio Grande, Rio Grande, Rua Eng. Alfredo Huch 475, Rio Grande, 96201-900, Brazil

^e Programa de Pós-Graduação em Enfermagem Fundação, Universidade Federal do Rio Grande, Rio Grande, Rua Eng.
Alfredo Huch 475, Rio Grande, 96201-900, Brazil

Received 1 July 2002; received in revised form 15 May 2003; accepted 15 June 2003

Abstract

The vasorelaxing activity of the aqueous extract of fish *Balistes capriscus* skin (AEBc) on mesenteric arterial bed (MAB) of rats was studied. The bolus injections of AEBc (bolus of 5.1, 10.2, 20.5, and 41.1 mg) significantly inhibited, in a concentration-dependent manner, the maximal contractile response induced by methoxamine (30 μ M) in MAB. The vasodilatation action of AEBc is not mediated through β -adrenoceptors or cyclo-oxygenase, since it was not affected by propranolol (20 μ M) or diclofenac sodium (3 μ M). The vasodilator response induced by subsequent addition of AEBc *Balistes capriscus* in bolus was significantly reduced in water infusion for endothelium removal. Treatment with an inhibitor of NO synthase (L-NAME, 10 μ M) decreased AEBc effect. The guanylate cyclase inhibitor methylene blue (MB, 100 μ M) had no significant effect on AEBc-induced vasodilatation. These results suggest that the vasorelaxing effect of AEBc is mediated by endothelium-dependent (NO/EDRF) and endothelium-independent neurally induced vasorelaxation from nonadrenergic and noncholinergic nerves (NO).

© 2003 Elsevier Ireland Ltd. All rights reserved.

Keywords: Nonadrenergic; Mesenteric arterial bed; Vasodilatation

1. Introduction

Investigation of biological, chemical, and ecological phenomena in the marine world contributes to a better understanding of marine habitats, and benefits the applied and basis research in pharmaceutical marine natural products (Konig and Wright, 1995), while research into the pharmacological properties of marine natural products may lead to promising candidates for new drugs and potentially active agents for clinical application. Several marine natural products are, in fact, currently in pre-clinical and clinical evaluation, while others show promising biological activities in in vivo and in vitro assays.

The sesterpenoide manoalide, for example, isolated from the sponge *Luffariella variabilis* in a program to search for

new anti-inflammatory compounds, proved to be a potent inhibitor of phospholipase C, (Konig and Wright, 1995).

In the city of Rio Grande, Southern of Brazil, the fish *Balistes capriscus* (tetradontiform, balistidae) has been used by fishermen in the treatment of respiratory disorders, such as asthma and bronchitis as water infusion of dried and powdered skin.

The present study was carried out to investigate the effects of aqueous extract of *Balistes capriscus* skin on isolated mesenteric arterial bed (MAB) precontracted with methoxamine for possible vascular relaxing activity.

2. Material and methods

2.1. Biological materials

Samples of the fish *Balistes capriscus* were collected during 1999 and 2000 along the coast of Rio Grande, RS

* Corresponding author. Tel.: +55-53-2338656;

fax: +55-53-2338680.

E-mail address: abaisch@octopus.furg.br (A.L. Muccillo-Baisch).

(southern Brazil) and identified by Dr. Manoel Haimovich from the Department of Oceanography of Fundação, Universidade Federal do Rio Grande (FURG), Rio Grande, Brazil. A voucher specimen has been deposited at the FURG fish collection (acquisition numbers 83.0245 and 83.0295). The animals were frozen at -18°C until used for the assays.

2.2. Extract

Fish skins were dried/toasted in a conventional oven, at the temperature of approximately 180°C . Coarsely powdered dried skin material (35 g) was extracted with 120 ml of bi-distilled water using a Soxhlet extractor, yielding, after lyophilization, 4.79 g of powder (13.7%). The extract was stored and dissolved in Krebs Ringer bicarbonate immediately before all tests were performed. All results are shown as milligram of lyophilized powder.

2.3. Animals

Male Wistar rats, weighing 419.88 ± 6.25 g ($n = 52$), were obtained from Animal House of the Fundação Universidade Federal do Rio Grande. The rats were housed in temperature-controlled rooms ($20\text{--}22^{\circ}\text{C}$), with a 12-h-light/12-h-dark photoperiod and $55 \pm 1\%$ relative humidity. Standard laboratory chow (Nuvital, Nuvital Nutrientes, Colombo, Paraná, Brazil) and drinking water were provided ad libitum.

2.4. Perfused mesenteric arterial vascular bed of the rat (MAB)

The MAB was removed from animals under brief ether anesthesia and perfused as described by Mc Gregor (1965), as modified by Silva et al. (1984). The superior mesenteric artery with its nerve plexus was carefully separated from the surrounding tissues at a point 2 cm distal to the aorta. A stainless steel cannula was inserted into the artery. The associated vascular bed was covered with cotton cloth moistened with Krebs Ringer bicarbonate and perfused by means of a peristaltic pump (Milan, Colombo, Brasil) at 5 ml/min with a similar solution. The composition of the perfusion solution was (mM): NaCl, 118; KCl, 4.7; CaCl_2 , 3.3; KH_2PO_4 , 1.2; MgSO_4 , 2.4; NaHCO_3 , 25; glucose, 10; EDTA, 0.03; ascorbic acid, 0.1. The Krebs Ringer bicarbonate solution was gassed with 95% O_2 and 5% CO_2 to obtain a pH of 7.2–7.4 and was maintained at 37°C .

Perfusion pressure was measured with a transducer (Hewlett-Packard, USA) on a side arm, just before the perfusing cannula, and was continuously recorded on a polygraph (Hewlett-Packard). Since the flow rate was constant throughout the whole experimental period, any pressure alteration reflected changes in vascular resistance. All

preparations were allowed to equilibrate for at least 30 min before the start of the experiments.

2.5. Vasodilatory responses

In order to evaluate the vasoactive effect of the extracts and perivascular nerve stimulation (PNS), the perfusion pressure of MAB was increased by infusing methoxamine ($30\ \mu\text{M}$). When a plateau was reached, the extract and all agonists were administered by bolus injection ($50\ \mu\text{M}$). Drugs and extract-induced relaxation (5.1, 10.2, 20.5, and 41.1 mg) was expressed as a percentage of the pressure induced by methoxamine ($30\ \mu\text{M}$).

In experiments with PNS two electrodes, one placed around the hypodermic needle used to cannulate the superior mesenteric artery, the other resting on the vasculature in a lower part of the vascular bed, were used to create transmural field stimulation (amplitude supra maximal of 60 V and pulse duration of 0.5 ms). Currents were applied at various frequencies and time using an electronic stimulator (HSE-D 7801-Hugstetten, Germany).

In order to block the vasoconstrictive action of perivascular sympathetic innervation, the MAB was perfused continuously with guanethidine ($5\ \mu\text{M}$), introduced after a period of equilibration. To evaluate vasodilating response, the tone of mesenteric vascular bed was raised by continuous perfusion with methoxamine ($30\ \mu\text{M}$), which was added to the perfusate, in order to maintain active tone of the mesenteric vasculature. The solution also contained guanethidine ($5\ \mu\text{M}$) to block norepinephrine release. The increased perfusion pressure was allowed to stabilize and the preparation was again subjected to PNS, applied for 30 s.

To study the influence of sensory neurones in the vasodilating responses to the extract or PNS, the preparations were perfused intra- and extra-luminally with capsaicin solution ($0.1\ \mu\text{M}$) for 20 min.

2.6. Drugs

The following drugs were used: acetylcholine chloride, N^{G} -nitro-L-arginine methyl ester (L-NAME) hydrochloride, methoxamine, papaverine, propranolol, salbutamol, guanethidine, capsaicin, and atropine from Sigma Chemical Co. (St. Louis, MO, USA), while sodium diclofenac (Voltaren[®] injection solution) was supplied by Novartis (São Paulo, Brazil). The stock solution of capsaicin was prepared in 70% ethanol; all other compounds were dissolved in distilled water.

2.7. Statistical analysis

Data are reported as mean \pm standard error of mean (S.E.M.). Statistical differences between means were evaluated using Student's *t* test for unpaired and paired observations. The level of significance was 95%.

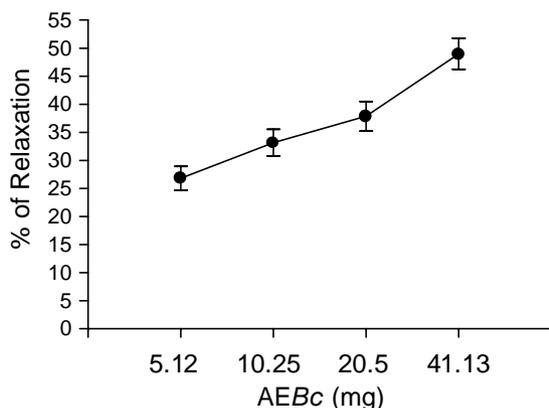


Fig. 1. Dose-response curve for the relaxation induced by different boluses of *Balistes capriscus* skin aqueous extract (AEBc) in the methoxamine-precontracted mesenteric arterial bed. Each data point on the graph represents the means \pm S.E.M. of 52 experiments.

3. Results

3.1. Vasodilatation in response to aqueous extract of *Balistes capriscus* skin (AEBc) in MAB precontracted with methoxamine

The basal perfusion pressure, of the MAB precontracted with methoxamine, was 19.7 ± 0.7 mm Hg ($n = 52$).

AEBc ($n = 52$) bolus injections (5.1, 10.2, 20.5, and 41.1 mg) significantly inhibited, in a concentration-dependent manner, the maximal contractile response induced by methoxamine (Fig. 1).

3.2. Effect of cyclooxygenase pathway inhibitor sodium diclofenac on changes in vasodilator responses to AEBc in mesenteric arterial bed precontracted by methoxamine

To exclude the involvement of prostaglandins in the extract-induced vasodilatation, the MAB was perfused with

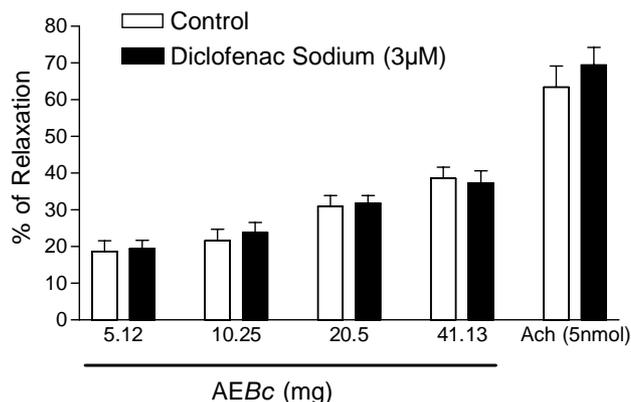


Fig. 2. Effect of sodium diclofenac ($3 \mu\text{M}$) on relaxation response to *Balistes capriscus* skin aqueous extract (AEBc) and acetylcholine (Ach) in methoxamine-precontracted mesenteric arterial bed. Relaxation is expressed as a percentage of the contraction induced by methoxamine. The values are means \pm S.E.M. ($n = 8$).

diclofenac sodium ($3 \mu\text{M}$) for 30 min. The treatment of preparations with the cyclooxygenase inhibitor ($n = 8$) did not modify AEBc responses (Fig. 2).

3.3. Effect of β -adrenergic pathway inhibitor on AEBc-induced vasodilatation in mesenteric arterial bed precontracted with methoxamine

To exclude the involvement of the β -adrenergic pathway on extract-induced vasodilatation, the MAB ($n = 6$) was perfused with propranolol ($20 \mu\text{M}$) for 30 min. The treatment of preparations with the inhibitor did not modify AEBc response (Fig. 3). On the other hand, the vasodilatory effect of salbutamol (8.0 nmol), a β -adrenergic agonist, was significantly modified, from $32.59 \pm 3.85\%$ to $14.33 \pm 4.97\%$ ($n = 6$) in the presence and absence of propranolol, respectively.

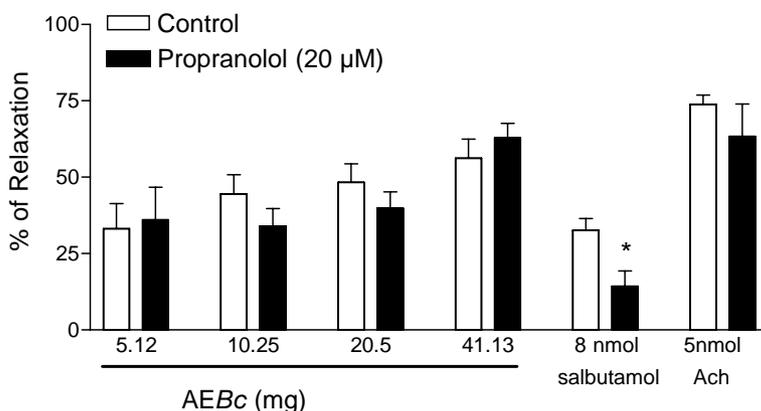


Fig. 3. Effects of propranolol ($20 \mu\text{M}$) on relaxation response to *Balistes capriscus* aqueous extract (AEBc), salbutamol and acetylcholine (Ach) in methoxamine-precontracted mesenteric arterial bed. Relaxation is expressed as a percentage of the contraction induced by methoxamine. The values are means \pm S.E.M. ($n = 6$). (*) Denotes value significantly different ($P < 0.05$) from the control.

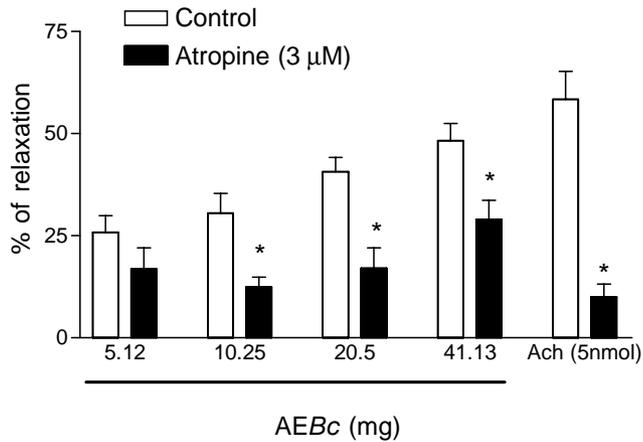


Fig. 4. Effects of atropine (3 μM) on relaxation response to *Balistes capriscus* skin aqueous extract (AEBc) and acetylcholine (Ach) in methoxamine-precontracted mesenteric arterial bed. Relaxation is expressed as a percentage of the contraction induced by methoxamine. The values are means ± S.E.M. ($n = 7$). (*) Denotes value significantly different ($P < 0.05$) from control.

3.4. Effect of muscarinic pathway inhibitor on AEBc and acetylcholine-induced vasodilatation in mesenteric arterial bed precontracted with methoxamine

To study the involvement of muscarinic pathway on extract-induced vasodilatation, the MAB was perfused with atropine for 30 min. As shown in Fig. 4, the extract-induced vasodilatation of the MAB were significantly inhibited ($P < 0.05$) by treatment with atropine (3 μM). The same procedure decreased the acetylcholine-induced vasodilatation from $58.38 \pm 6.82\%$ to $10.0 \pm 3.1\%$.

3.5. Effect of capsaicin treatment on changes in perfusion pressure induced by AEBc and perivascular nerve stimulation in MAB precontracted with methoxamine

Capsaicin (0.1 μM) pretreatment for 20 min prevented the vasodilatation induced by aqueous extract of *Balistes*

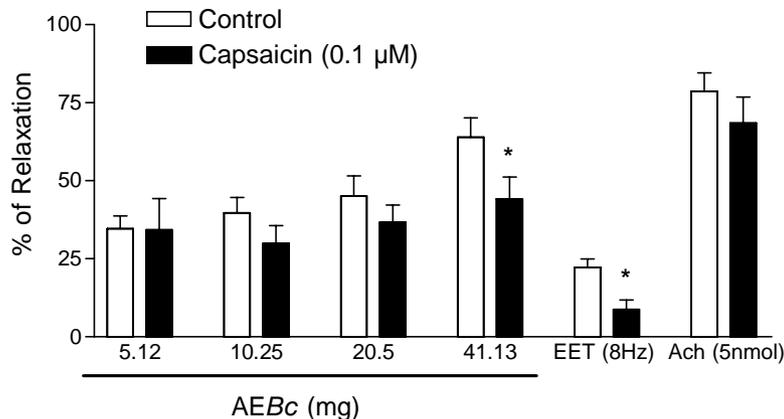


Fig. 5. Effects of capsaicin (0.1 μM) on relaxation response to *Balistes capriscus* skin aqueous extract (AEBc) and PNS-induced vasodilatation in methoxamine-precontracted mesenteric arterial bed. Relaxation is expressed as a percentage of the contraction induced by methoxamine. The values are means ± S.E.M. ($n = 8$). (*) Denotes value significantly different ($P < 0.05$) from control.

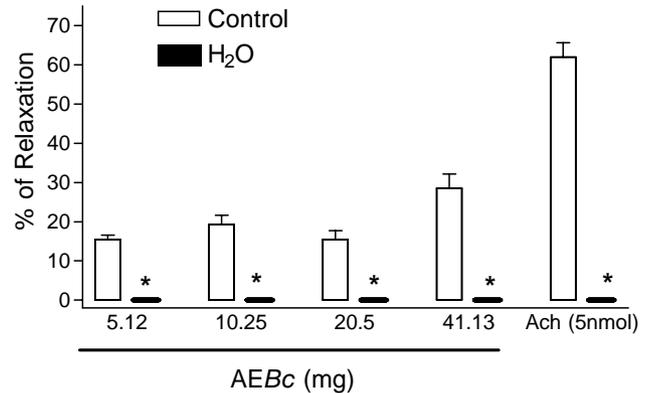


Fig. 6. Effects of endothelium removal on relaxation response to *Balistes capriscus* skin aqueous extract (AEBc) and acetylcholine (Ach) in methoxamine-precontracted mesenteric arterial bed. Relaxation is expressed as a percentage of the contraction induced by methoxamine. The values are means ± S.E.M. ($n = 8$). (*) Denotes value significantly different ($P < 0.05$) from control.

capriscus, but only for the bolus of 41.3 mg, from $63.97 \pm 6.15\%$ to $44.19 \pm 7.0\%$ (Fig. 5).

The response of MAB to PNS in the presence of capsaicin was inhibited from $22.28 \pm 2.66\%$ to $8.78 \pm 2.97\%$ (Fig. 5).

3.6. Effect of endothelium removal on changes in perfusion pressure induced by AEBc and acetylcholine in MAB precontracted with methoxamine

To exclude the involvement of endothelium on extract-induced vasodilatation, the MAB was perfused with distilled water for 10 min for endothelium removal (Criscione et al., 1984). In the intact MAB ($n = 8$), contracted with methoxamine (30 μM), aqueous extractable fraction caused immediate relaxation. After the endothelium was removed the AEBc-induced vasodilatation was significantly changed (Fig. 6). The same procedure decreased the acetylcholine-induced vasodilatation (5 nmol, bolus) from $62.0 \pm 3.65\%$ to zero.

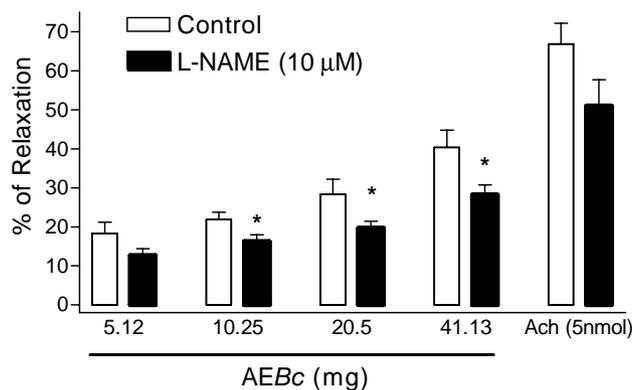


Fig. 7. Effects of N^G -nitro-L-arginine methyl ester (L-NAME, 10 μ M) on relaxation response to *Balistes capriscus* skin aqueous extract (AEBc) and acetylcholine (Ach) in methoxamine-precontracted mesenteric arterial bed. Relaxation is expressed as a percentage of the contraction induced by methoxamine. The values are means \pm S.E.M. ($n = 8$). (*) Denotes value significantly different ($P < 0.05$) from control.

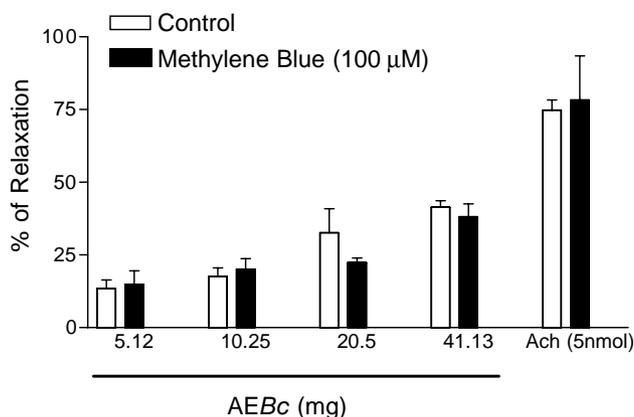


Fig. 8. Effects of methylene blue (100 μ M) on relaxation response to *Balistes capriscus* skin aqueous extract (AEBc) and acetylcholine (Ach) in methoxamine-precontracted mesenteric arterial bed. Relaxation is expressed as a percentage of the contraction induced by methoxamine. The values are means \pm S.E.M. ($n = 7$).

3.7. Effect of endothelium-derived NO-cyclic GMP pathway on vasodilating responses to AEBc in mesenteric arterial bed-precontracted with methoxamine

As shown in Fig. 7, the treatment of preparation with an inhibitor of NO synthase L-NAME (10 μ M) significantly reduced the AEBc-induced vasodilatation.

The effect of guanylate cyclase inhibitor methylene blue (MB, 100 μ M) on the vasodilatation induced by AEBc was also examined. The treatment of preparations with the inhibitor did not significantly modify the AEBc response (Fig. 8).

4. Discussion

The results of the present study demonstrate, for the first time, that the skin of *Balistes capriscus*, used in popular

medicine in Southern Brazil for treatment of respiratory disorders, produced a dose-dependent and reversible inhibition of methoxamine-induced vascular tone.

The vasorelaxant action of the extract does not appear to involve activation of β -adrenoceptors, since incubation of the preparations with propranolol failed to affect its vasorelaxation action.

The relaxation induced by the extract was not affected by sodium diclofenac, which indicates that it is not indirectly mediated by the release of prostanoid metabolite(s) derived from the cyclooxygenase pathway of arachidonic acid.

As is well known, the vasorelaxing action of acetylcholine requires the presence of an intact endothelium. The binding of acetylcholine to muscarinic receptors on endothelial cells trigger the release of a potent vasodilator called endothelium-derived relaxing factor (EDRF), discovered by Furchgott and Zawadzki (1980). The EDRF, as nitric oxide (NO), is synthesized from guanidino groups of L-arginine (Palmer et al., 1987) and has been reported to stimulate the production of cGMP, which induces a vasorelaxation in smooth muscle cells (Winquist et al., 1984).

The treatment of the preparations with atropine, at concentration where Ach-mediated vasorelaxation was antagonized (Muccillo Baisch et al., 1998), prevents a great part of the vasorelaxant action of the extract of *Balistes capriscus* skin in MAB. In addition, when the MAB was perfused with distilled water, the vasorelaxant actions of acetylcholine and of the extract were greatly reduced. As exposure to distilled water destroys the endothelial cells (Criscione et al., 1984), these results confirm that in MAB of rats, both acetylcholine (Muccillo Baisch et al., 1998) and the extract-induced vasodilatation are mediated by the release of endothelium-derived substances. Taken together, such results suggest that the vasorelaxant actions of the extract are mediated jointly by endothelium-dependent cholinergic mechanism.

The vasorelaxant effects of endothelium-dependent substances could be inhibited by several L-arginine analogues such as *N*-monomethyl-L-arginine (L-NMMA) and N^G -nitro-L-arginine methyl ester (L-NAME) (Palmer et al., 1987; Rees et al., 1989 and Moore et al., 1990). In the present study the aqueous extract of *Balistes capriscus* dried skin caused endothelium-dependent relaxation, which was reversed by L-NAME, at a concentration which Ach-mediated vasorelaxation was antagonized. This suggests that certain compounds present in the skin extract may cause relaxation by increasing NO or other related compounds.

The endothelium-dependent vasorelaxants, such as acetylcholine, could also release endothelium-derived hyperpolarizing factor (EDHF) from the endothelium, which induces vasorelaxation through membrane hyperpolarization (Adeagbo and Triggle, 1993). The possible existence of a novel endothelium-derived relaxing factor in the endothelium of rat MAB was examined in the study of Kamata et al. (1996). The authors suggested that one or more EDRF must exist, other than NO or EDHF. The novel EDRF may relax

the MAB through production of cAMP, but not cGMP. This suggestion could support the finding that the vasorelaxant effect induced by AEBc was not inhibited by methylene blue. However, methylene blue is not a potent guanylate cyclase inhibitor, and has also been shown to inhibit NO's activity (Mayer et al., 1993; Lou et al., 1995).

In the MAB, the relaxation induced by PNS has been determined to be mediated by the activation of capsaicin-sensitive sensory fibers which releases neuropeptides locally, with a potent vasorelaxation action on intestinal blood flow (Manzini and Perreti, 1988). Thus, the present investigation has demonstrated the presence of a nonadrenergic, noncholinergic vasodilator innervation in rat mesenteric arteries.

In the present study, the vasodilatation induced by aqueous extract from *Balistes capriscus* skin was reduced in the presence of capsaicin, a pungent ingredient in chili pepper (*Capsicum* sp.), which has been used to damage selectively the subpopulation of sensory neurons and to deplete neurotransmitter, such as substance P and CGRP (Holzer, 1988). This suggests that the *Balistes capriscus* skin-induced MAB vasodilatation is mediated by primary sensory nerve fibers. Consequently these data show that in the MAB, the vasodilatation induced by the aqueous extract of *Balistes capriscus* skin is mediated by endothelium-dependent mechanism mediated by EDHF or NO and endothelium-independent neurally induced relaxation, associated with NO from a nonadrenergic noncholinergic nerves.

Pharmacological and chemical studies are currently in progress to isolate and characterize the active compounds present in the fish skin, and also to investigate the possible mechanism of action.

Acknowledgements

The authors are grateful to Prof. Dr. Manoel Haimovich, for the donation and identification of biological material, Mr. Robaldo, M.Sc. for help in the taxonomic identification of the fish, Walquíria Lopes for technical assistance, Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) for Mayeve Grando and Carla Krueel financial assistance, Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) for M.Sc. fellowship to Lisandra Cavalli, and Fundação de Amparo a Pesquisa do Rio Grande do Sul (FAPERGS) for Paula Lidiane Possette's financial assistance. Thanks are also due to Dr. Euclides dos Santos F^o and Dr. Luiz Eduardo Maia Nery for assisting in the revision of the manuscript.

References

- Adeagbo, A.S.O., Triggle, C.R., 1993. Varying extracellular [K⁺]: a functional approach to separating EDHF- and EDNO-related mechanisms in perfused rat mesenteric arterial bed. *Journal of Cardiovascular Pharmacology* 21, 423–429.
- Criscione, L., Müller, K., Prescott, M.F., 1984. Endothelial cells loss enhances the pressor response in resistance vessels. *Journal of Hypertension* 2, 441–444.
- Furchgott, R.F., Zawadzki, J.V., 1980. The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. *Nature* 299, 373–376.
- Holzer, P., 1988. Local effector functions of capsaicin-sensitive sensory nerve endings: involvement of tachykinins CGRP and other neuropeptides. *Neuroscience* 24, 739–768.
- Kamata, K., Numazawa, T., Kasuya, Y., 1996. Characteristics of vasodilatation induced by acetylcholine and platelet-activating factors in the rat mesenteric arterial bed. *European Journal of Pharmacology* 298, 129–136.
- Konig, G.M., Wright, A.D., 1995. Marine natural products research: current directions and future potential. *Planta Medica* 62, 193–211.
- Lou, D., Das, S., Vincent, S.R., 1995. Effects of methylene blue and LY83563 on neuronal nitric oxide synthase and NADPH-diaphorase. *European Journal of Pharmacology* 290, 247–251.
- Manzini, S., Perreti, F., 1988. Vascular effects of capsaicin in isolated perfused rat mesenteric bed. *European Journal of Pharmacology* 148, 153–159.
- Mayer, B., Brunner, F., Schmidt, K., 1993. Inhibition of nitric oxide synthesis by methylene blue. *Biochemical Pharmacology* 45, 367–374.
- Mc Gregor, D.D., 1965. The effect of sympathetic nerve stimulation on vasoconstrictor responses in perfused mesenteric blood vessel of the rat. *Journal of Physiology* 177, 21–30.
- Moore, P.K., al-Swayeh, O.A., Chong, N.W.S., Evans, R.A., Gibson, A., 1990. L-N^G-nitro arginine (L-NOARG), a novel, L-arginine-reversible inhibitor of endothelium-dependent vasodilatation in vitro. *British Journal of Pharmacology* 99, 408–412.
- Muccillo Baisch, A.L., Johnston, K.B., Paganini Stein, F.L., 1998. Endothelium-dependent vasorelaxing activity of aqueous extract of *Ilex paraguariensis* on mesenteric arterial bed of rats. *Journal of Ethnopharmacology* 60, 133–139.
- Palmer, R.M.J., Ferridge, A.G., Mocada, S., 1987. Nitric oxide release accounts for the biological activity of endothelium-derived relaxing factor. *Nature* 327, 524–526.
- Rees, D.D., Palmer, R.M.J., Moncada, S., 1989. Role of endothelium-derived nitric oxide in the regulation of blood pressure. *Proceeding of the National Academy of Science (USA)* 86, 3375–3378.
- Silva, F.A., Muccillo Baisch, A.L., Almeida, T.U., Castro, J.A., 1984. A vasodilator action of nicotine on the isolated superior mesenteric artery. In: *Proceedings II Congress of the Brazilian Society of Pharmacology and Experimental Therapeutics*. *Brazilian Journal of Medical and Biological Research* 17, 510.
- Winquist, R.J., Bunting, P.B., Baskin, E.P., Wallace, A.A., 1984. Decreased endothelium-dependent relaxation in New Zealand genetic hypertensive rats. *Journal of Hypertension* 2, 536–541.